

## DC Medium with BCIG (NCM0112)

### Intended Use

DC Medium with BCIG is used in the chromogenic differentiation of *E. coli* from other coliforms in water samples using the membrane filtration method in a laboratory setting. DC Medium with BCIG is not intended for use in the diagnosis of disease or other conditions in humans.

### Description

DC (Differential Coliform) Medium with BCIG is a selective and differential medium for the presumptive identification of *E. coli*. This medium is enhanced by the addition of a chromogenic agent, BCIG, 5-bromo-4-chloro-3-indolyl- $\beta$ -D-glucuronide to detect glucuronidase activity. The presence of the enzyme  $\beta$ -D-glucuronidase differentiates most *E. coli* spp. from other coliforms, and is the same enzyme used in the MUG reaction. BCIG reacts slightly differently, and when released into the medium is insoluble, accumulating within the cell, imparting a blue to purple color to presumptive *E. coli* colonies.

### Typical Formulation

Lactose	10.0 g/L
Tryptose	10.0 g/L
Yeast Extract	3.0 g/L
Sodium Chloride	5.0 g/L
Proteose Peptone	5.0 g/L
Bile Salts	1.5 g/L
BCIG	0.2 g/L
Neutral Red	0.08 g/L
Agar	15.0 g/L

Final pH: 7.2  $\pm$  0.2 at 25°C

Formula may be adjusted and/or supplemented as required to meet performance specifications.

### Supplement

Cefsulodin solution, 10 mg/mL

### Precaution

Refer to SDS

### Preparation

1. Suspend 49.8 g of the medium in one liter of purified water.
2. Heat with frequent agitation and boil for one minute to completely dissolve the medium.
3. DO NOT AUTOCLAVE.
4. Cool to 45 - 50°C.
5. Aseptically add 1.2 mL of a sterile 10 mg/mL solution of Cefsulodin and mix well.

### Quality Control Specifications

**Dehydrated Appearance:** Powder is homogeneous, free flowing, and beige to grayish pink with blue specs.

**Prepared Appearance:** Prepared medium is clear to trace hazy and red.

**Expected Cultural Response:** Cultural response on DC Medium with BCIG using the Membrane Filtration Method. Cultures were incubated at 35  $\pm$  2°C and examined for growth after 22 - 24 hours.

# Technical Specification Sheet



Microorganism	Approx. Inoculum (CFU)	Expected Results	Expected Reactions
<i>Escherichia coli</i> ATCC® 25922	10 - 100	Growth	Blue / purple colonies
<i>Citrobacter freundii</i> ATCC® 8090	10 - 100	Growth	Pink colonies or white w/ pink center colonies
<i>Proteus mirabilis</i> ATCC® 12453	10 - 100	Growth	Small pale salmon colonies
<i>Staphylococcus aureus</i> ATCC® 25923	1000	Completely Inhibited	---

The organisms listed are the minimum that should be used for quality control testing.

## Test Procedure

### Membrane Filtration Technique

1. Filter appropriate aliquot of the sample(s) to be tested through a cellulose membrane.
2. Transfer the membrane to a prepared plate of DC Medium with BCIG, supplemented with Cefsulodin.
3. Incubate at 35°C for 22 – 24 hours.
4. Examine plates for growth of *E. coli* colonies.

For a complete discussion on Membrane Filtration Procedures, refer to appropriate references.

## Results

*E. coli* colonies are blue/purple. The total number of *E. coli* per gram can be calculated by multiplying the blue/purple colonies by the dilution factor. The number of presumptive *E. coli* is obtained by multiplying the number of blue/purple colonies by the dilution factor.

## Expiration

Refer to expiration date stamped on the container. The dehydrated medium should be discarded if not free flowing, or if the appearance has changed from the original color. Expiry applies to medium in its intact container when stored as directed.

## Limitation of the Procedure

Due to varying nutritional requirements, some strains may be encountered that grow poorly or fail to grow.

## Storage

Store dehydrated culture media at 2-30°C away from direct sunlight. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light by keeping container tightly closed.

## References

1. Feng, P. C. S., and P. A. Hartmann. 1982. Appl. Environ. Microbiol. 43:1320-1329.
2. [www.fda.gov/Food/ScienceResearch/LaboratoryMethods/BacteriologicalAnalyticalmanualBAM/default.htm](http://www.fda.gov/Food/ScienceResearch/LaboratoryMethods/BacteriologicalAnalyticalmanualBAM/default.htm).
3. Vanderzant, C., and D. F. Splittstoesser (eds.). 2015. Compendium of methods for the microbiological examination of foods, 4<sup>th</sup> ed. American Public Health Association, Washington, D.C.

