CERTIFICATION

AOAC® *Performance Tested*SM

Certificate No.

071601

The AOAC Research Institute hereby certifies the performance of the test kit known as:

ANSR® for *Campylobacter*

manufactured by

Neogen Corporation
620 Lesher Place
Lansing, Michigan 48912
USA

This method has been evaluated in the AOAC® *Performance Tested Methods*SM Program and found to perform as stated by the manufacturer contingent to the comments contained in the manuscript. This certificate means that an AOAC® Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC *Performance Tested*SM certification mark along with the statement - "THIS METHOD'S PERFORMANCE WAS REVIEWED BY AOAC RESEARCH INSTITUTE AND WAS FOUND TO PERFORM TO THE MANUFACTURER'S SPECIFICATIONS" - on the above mentioned method for a period of one calendar year from the date of this certificate (November 24, 2019 – December 31, 2020). Renewal may be granted at the end of one year under the rules stated in the licensing agreement.

Scott Coates, Senior Director
Signature for AOAC Research Institute

November 24, 2019
Date

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SUBMITTING COMPANY
Neogen Corporation
620 Lesher Place
Lansing, MI 48912

KIT NAME(S)
ANSR® for *Campylobacter*

CATALOG NUMBERS
9872

INDEPENDENT LABORATORY
Q Laboratories, Inc.
1400 Harrison Avenue
Cincinnati, OH 45214
USA

AOAC EXPERTS AND PEER REVIEWERS
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APPLICABILITY OF METHOD
Target organism – *Campylobacter jejuni*, *Campylobacter lari*, and *Campylobacter coli*

Matrices – chicken carcass rinse and turkey carcass sponge

Performance claims - As determined by probability of detection analysis, the ANSR® for *Campylobacter* method performance is equivalent to the U.S. Department of Agriculture Food Safety and Inspection Service Microbiology Laboratory Guidebook (USDA-FSIS/MLG), Ch. 41.04, Isolation and Identification of *Campylobacter jejuni/coli/lari* from Poultry Rinse, Sponge and Raw Product Samples (2).

REFERENCE METHOD
USDA-FSIS (2016) *Microbiology Laboratory Guidebook*, Chapter 41.04, Isolation and Identification of *Campylobacter jejuni/coli/lari* from Poultry Rinse, Sponge and Raw Product Samples (Accessed February, 2016)
http://www.fsis.usda.gov/wps/wcm/connect/0273bc3d-2363-45b3-befb-1190c25f3c8b/MLG-41.pdf?MOD=AJPERES (2)

ORIGINAL CERTIFICATION DATE
July 02, 2016

CERTIFICATION RENEWAL RECORD
Renewed annually through December 2020

METHOD MODIFICATION RECORD
1. **January 2018 Level 1**
2. **December 2018 Level 1**
3. **November 2019 Level 1**

SUMMARY OF MODIFICATION
1. **Editorial changes**
2. **Editorial changes to add equipment, temperature ranges, correct issues in insert**
3. **Editorial changes**

Under this AOAC® *Performance Tested*℠ License Number, 071601 this method is distributed by:
NONE

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NONE

PRINCIPLE OF THE METHOD (1)
ANSR® for *Campylobacter* is an isothermal, amplified nucleic acid assay, and the method is based on NEAR™ technology. Briefly, the amplification reaction is preceded by the reverse transcription of 16S ribosomal RNA. Target complementary DNA is amplified through a polymerization mechanism from the ends of nicks created in double-stranded DNA by specific endonuclease action. The nicked DNA is then amplified using specific templates and a DNA polymerase. Finally, amplified target sequences are detected using fluorescent molecular beacon probes.
Samples are analyzed following a 20–24 h enrichment at 42°C ± 1°C in ANSR for *Campylobacter* enrichment broth in either a microaerobic or aerobic atmosphere. Then, a two-stage lysis reaction is performed on the enriched sample, first at 37 ± 2°C for 10 min, then at 80 ± 2°C for 20 min. Next, a portion of the lysed sample is transferred to a strip tube containing lyophilized ANSR reagents. The tubes are sealed and incubated at 56 ± 1°C on the ANSR reader. Results are generated by the reader and displayed in the ANSR software within 20 minutes as positive, negative or invalid. Invalid assay results must be repeated, while positive results may be confirmed with enrichment cultures following standard procedures (7). Campy-Cefex Agar is recommended as the plating media. Each tube of ANSR reagents contains an internal positive control, which ensures that the reagents are functioning properly. The ANSR® for *Campylobacter* test is designed for use by personnel with appropriate training in microbiology. Training in the use of the ANSR test system is available through Neogen.

DISCUSSION OF THE VALIDATION STUDY (1)
The results of this study provide evidence that the overall performance of the ANSR for *Campylobacter* assay is equivalent to the USDA-FSIS/MLG reference method. Based on internal and independent laboratory studies for chicken carcass rinse and turkey carcass sponge, ANSR for *Campylobacter* is an effective alternative for detection of *C. jejuni*, *C. lari*, and C. *coli* after 20–24 h of enrichment in microaerobic or aerobic atmosphere. Inclusivity testing produced 100% positive results in testing of 50 *C. jejuni*, *C. lari*, and *C. coli* strains. With all 31 exclusivity strains testing negative, including non-target *Campylobacter* species, the ANSR for *Campylobacter* test shows 100% target specificity. Robustness results provide evidence that the assay can withstand modest procedural changes simultaneously and still produce accurate results.
Finally, ANSR for *Campylobacter* offers the efficiency of a 20 h enrichment step, coupled with the flexibility of up to a 24 h single-step enrichment in microaerobic or aerobic atmosphere. Furthermore, the ANSR for *Campylobacter* method offers the advantages of minimal labor and assay hardware requirements and results within 50 minutes following sample enrichment.

Table 1. Results of inclusivity testing for the ANSR for *Campylobacter* test using ANSR for *Campylobacter* enrichment media with incubation in microaerobic or aerobic atmosphere (1)

Strain No.	*ATCC[a] No.*	*Organism*	*Source (if known)*	*Origin (if known)*	*ANSR Result*	
					Microaerobic[b]	*Aerobic[c]*
16	33559	*Campylobacter coli*		Pig feces	Positive	Positive
	BAA-1061	*Campylobacter coli*		Chicken carcass	Positive	Positive
669		*Campylobacter coli*	Neogen Corp.[d]		Positive	Positive
2728		*Campylobacter coli*	Neogen Corp.		Positive	Positive
2739		*Campylobacter coli*	Neogen Corp.		Positive	Positive
2749		*Campylobacter coli*	Neogen Corp.		Positive	Positive
2751		*Campylobacter coli*	Neogen Corp.		Positive	Positive
2755		*Campylobacter coli*	Neogen Corp.		Positive	Positive
3051		*Campylobacter coli*	Neogen Corp.		Positive	Positive
2757		*Campylobacter coli*	Neogen Corp.		Positive	Positive
	BAA-1062	*Campylobacter jejuni*		Chicken carcass	Positive	Positive
23	29428	*Campylobacter jejuni*		Human child feces	Positive	Positive
22	33560	*Campylobacter jejuni*		Bovine feces	Positive	Positive
3581	43429	*Campylobacter jejuni*		Human feces	Positive	Positive
3582	43430	*Campylobacter jejuni*		Animal feces	Positive	Positive
3583	43431	*Campylobacter jejuni*		Human feces	Positive	Positive
3584	43433	*Campylobacter jejuni*		Human feces	Positive	Positive
3585	43438	*Campylobacter jejuni*		Human feces	Positive	Positive
3586	43442	*Campylobacter jejuni*		Human feces	Positive	Positive
2887		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2888		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2889		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2890		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2891		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2892		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2937		*Campylobacter jejuni*			Positive	Positive
2938		*Campylobacter jejuni*			Positive	Positive
2940		*Campylobacter jejuni*			Positive	Positive
2740		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2741		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2742		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
2743		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3050		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3052		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3054		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3225		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3226		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3227		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3228		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3229		*Campylobacter jejuni*	Neogen Corp.		Positive	Positive
3231		*Campylobacter jejuni*	Evanston hospital[e]	Stool	Positive	Positive
3232		*Campylobacter jejuni*	Evanston hospital	Stool	Positive	Positive
3255		*Campylobacter jejuni*	Evanston hospital	Stool	Positive	Positive
	35221	*Campylobacter lari*		Herring gull cloacal swab	Positive	Positive
24	35223	*Campylobacter lari*			Positive	Positive
801		*Campylobacter lari*	Neogen Corp.		Positive	Positive
2759		*Campylobacter lari*	Neogen Corp.		Positive	Positive
	43675	*Campylobacter lari*		Human feces	Positive	Positive
	35222	*Campylobacter lari*		Animal feces	Positive	Positive
	BAA-1060	*Campylobacter lari*	CDC[f]	Human feces	Positive	Positive

[a]American Type Culture Collection, Manassas, VI.

[b]Microaerobic is incubation in 85% nitrogen, 10% carbon dioxide, 5% oxygen, 42 ± 1°C for 22 ± 2 h.

[c]Aerobic is incubation in normal atmosphere at 42° ± 1°C for 22 ± 2 h.

[d]Neogen Corporation, Lansing, MI.

[e]Evanston hospital, Evanston, IL.

[f]Centers for Disease Control and Prevention, Atlanta, GA.

Table 2. Results of exclusivity testing for the ANSR for *Campylobacter* test (1)

Strain No.	*ATCC[a] No.*	*Organism*	*Source (if known)*	*Origin (if known)*	*ANSR Result*
2	19606	*Acinetobacter calcoaceticus*			Negative
3362	15468	*Aeromonas caviae*			Negative
17	43157	*Arcobacter cryaerophilus*		Eye of aborted porcine fetus	Negative
803	8186	*Bacillus parabrevis*			Negative
804	23059	*Bacillus subtilis*		Soil	Negative
10	23745	*Bacteroides fragilis*		Pleural Fluid	Negative
308	14686	*Bergeriella denitrificans*		Oral mucosa of guinea pig	Negative
807	33237	*Campylobacter concisus*		Human gingival sulcus	Negative
2224	35224	*Campylobacter curvus*		Human jaw abscess	Negative
802	33709	*Campylobacter sputorum* biovar *faecalis*		Sheep feces	Negative
19	33561	*Campylobacter fetus ss venerealis*		Human blood	Negative
2761	35217	*Campylobacter hyointestinalis*		Swine intestine	Negative
718		*Campylobacter rectus*			Negative
2913		*Campylobacter upsaliensis*	CDC[b]	stool	Negative
	10231	*Candida albicans*			Negative
	6879	*Citrobacter freundii*		Milk set for cottage cheese	Negative
	27156	*Citrobacter koseri*			Negative
43	9689	*Clostridium difficile*			Negative
2791		*Escherichia coli*	Sani-Pure Lab[c]	chocolate sprinkles, powder	Negative
1502	29939	*Klebsiella pneumoniae*			Negative
313	27633	*Neisseria gonorrhoeae*		Patient with gonorrhea	Negative
2197	14029	*Plesiomonas shigelloides*			Negative
797	25933	*Proteus mirabilis*		Human vagina	Negative
372	27970	*Providencia alcalifaciens*	CDC	Feces	Negative
682	33531	*Raoultella planticola*		Radish Root	Negative
799	13314	*Salmonella enterica* ssp. *Arizonae*			Negative
	9207	*Shigella boydii*			Negative
798	12022	*Shigella flexneri*			Negative
399	12600	*Staphylococcus aureus*		Plueral fluid	Negative
3326		*Vibrio vulnificus*	Gulf Coast Research Lab[d]	Oyster	Negative
418	35236	*Yersinia aldovae*		Drinking water	Negative

[a]American Type Culture Collection, Manassas, VI.
[b]Centers for Disease Control and Prevention, Atlanta, GA.
[c]Sani-Pure Lab, Saddle Brook, NJ
[d]Gulf Coast Research Lab, Dauphin Island, AL

Table 3. Probability of detection calculations for ANSR for *Campylobacter* presumptive and confirmed results after 20 h microaerobic or aerobic enrichment of naturally-contaminated samples (1)

Sample type	ANSR Incubation Condition[a]	N[b]	ANSR presumptive result			ANSR confirmed result			$dPOD_{CP}$[f]	95% CI[g]
			X[c]	POD_{CP}[d]	95% CI	X	POD_{CC}[e]	95% CI		
Chicken Rinse Sample Set 1	Aerobic	20	14	0.70	0.48, 0.85	14	0.70	0.48, 0.85	0.00	-0.27, 0.27
	Microaerobic	20	14	0.70	0.48, 0.85	14	0.70	0.48, 0.85	0.00	-0.27, 0.27
Chicken Rinse Sample Set 2	Aerobic	20	15	0.75	0.53, 0.88	15	0.75	0.53, 0.88	0.00	-0.26, 0.26
	Microaerobic	20	11	0.55	0.34, 0.74	11	0.55	0.34, 0.74	0.00	-0.28, 0.28
Chicken Rinse Sample Set 1[h]	Aerobic	20	6	0.30	0.15, 0.52	5	0.25	0.11, 0.47	0.05	-0.22, 0.31
	Microaerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
Chicken Rinse Sample Set 2[h]	Aerobic	20	6	0.30	0.15, 0.52	6	0.30	0.15, 0.52	0.00	-0.27, 0.27
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26
Turkey Sponge Sample Set 1	Aerobic	20	20	1.00	0.84, 1.00	20	1.00	0.84, 1.00	0.00	-0.16, 0.16
	Microaerobic	20	19	0.95	0.76, 1.00	19	0.95	0.76, 1.00	0.00	-0.19, 0.19
Turkey Sponge Sample Set 2	Aerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26

[a] The enrichment condition for the ANSR method (aerobic = 42 ± 1.0°C in normal atmosphere; microaerobic = 42 ± 1.0°C in microaerobic atmosphere, 85% nitrogen, 10% carbon dioxide, 5% oxygen).
[b] N = Number of test portions.
[c] X = Number of positive test portions.
[d] POD_{CP} = Probability of detection for candidate method presumptive positive outcomes.
[e] POD_{CC} = Probability of detection for candidate method presumptive positive outcomes confirmed positive.
[f] $dPOD_{CP}$ = Difference between the candidate method presumptive and candidate method confirmed POD values.
[g] 95% CI = If the confidence interval of a dPOD does not contain zero, then the difference is statistically significant at the 5% level.
[h] Trial performed by independent laboratory.

Table 4. Probability of detection calculations for ANSR for *Campylobacter* presumptive and confirmed results after 24 h microaerobic or aerobic enrichment of naturally-contaminated samples (1)

Sample Type	ANSR Incubation Condition[a]	N[b]	ANSR presumptive result			ANSR confirmed result			$dPOD_{CP}$[f]	95% CI[g]
			X[c]	POD_{CP}[d]	95% CI	X	POD_{CC}[e]	95% CI		
Chicken Rinse Sample Set 1	Aerobic	20	14	0.70	0.48, 0.85	14	0.70	0.48, 0.85	0.00	-0.27, 0.27
	Microaerobic	20	14	0.70	0.48, 0.85	14	0.70	0.48, 0.85	0.00	-0.27, 0.27
Chicken Rinse Sample Set 2	Aerobic	20	15	0.75	0.53, 0.88	15	0.75	0.53, 0.88	0.00	-0.26, 0.26
	Microaerobic	20	11	0.55	0.34, 0.74	11	0.55	0.34, 0.74	0.00	-0.28, 0.28
Chicken Rinse Sample Set 1[h]	Aerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26
	Microaerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
Chicken Rinse Sample Set 2[h]	Aerobic	20	6	0.30	0.15, 0.52	6	0.30	0.15, 0.52	0.00	-0.27, 0.27
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26
Turkey Sponge Sample Set 1	Aerobic	20	20	1.00	0.84, 1.00	20	1.00	0.84, 1.00	0.00	-0.16, 0.16
	Microaerobic	20	19	0.95	0.76, 1.00	19	0.95	0.76, 1.00	0.00	-0.19, 0.19
Turkey Sponge Sample Set 2	Aerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26

[a] The enrichment condition for the ANSR method (aerobic = 42 ± 1°C in normal atmosphere; microaerobic = 42 ± 1°C in microaerobic atmosphere, 85% nitrogen, 10% carbon dioxide, 5% oxygen).
[b] N = Number of test portions.
[c] X = Number of positive test portions.
[d] POD_{CP} = Probability of detection for candidate method presumptive positive outcomes.
[e] POD_{CC} = Probability of detection for candidate method presumptive positive outcomes confirmed positive.
[f] $dPOD_{CP}$ = Difference between the candidate method presumptive and candidate method confirmed POD values.
[g] 95% CI = If the confidence interval of a dPOD does not contain zero, then the difference is statistically significant at the 5% level.
[h] Trial performed by independent laboratory.

Table 5. Probability of detection calculations for ANSR for *Campylobacter* confirmed and reference method results after 20 h microaerobic or aerobic enrichment of naturally-contaminated samples (1)

Sample Type	ANSR Incubation Condition[a]	N[b]	ANSR confirmed result			Reference method result			$dPOD_C$[f]	95% CI[g]
			X[c]	POD_C[d]	95% CI	X	POD_R[e]	95% CI		
Chicken Rinse Sample Set 1	Aerobic	20	14	0.70	0.48, 0.85	17	0.85	0.64, 0.95	-0.15	-0.39, 0.11
	Microaerobic	20	14	0.70	0.48, 0.85	17	0.85	0.64, 0.95	-0.15	-0.39, 0.11
Chicken Rinse Sample Set 2	Aerobic	20	15	0.75	0.53, 0.88	16	0.80	0.58, 0.92	-0.05	-0.30, 0.21
	Microaerobic	20	11	0.55	0.34, 0.74	11	0.55	0.34, 0.74	0.00	-0.28, 0.28
Chicken Rinse Sample Set 1[h]	Aerobic	20	5	0.25	0.11, 0.47	7	0.35	0.18, 0.57	-0.10	-0.36, 0.18
	Microaerobic	20	7	0.35	0.18, 0.57	6	0.30	0.15, 0.52	0.05	-0.23, 0.32
Chicken Rinse Sample Set 2[h]	Aerobic	20	6	0.30	0.15, 0.52	8	0.40	0.22, 0.61	-0.10	-0.36, 0.18
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26
Turkey Sponge Sample Set 1	Aerobic	20	20	1.00	0.84, 1.00	20	1.00	0.84, 1.00	0.00	-0.16, 0.16
	Microaerobic	20	19	0.95	0.76, 1.00	20	1.00	0.84, 1.00	-0.05	-0.24, 0.12
Turkey Sponge Sample Set 2	Aerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
	Microaerobic	20	5	0.25	0.11, 0.47	7	0.35	0.18, 0.57	-0.10	-0.36, 0.18

[a] The enrichment condition for the ANSR method (aerobic = 42 ± 1°C in normal atmosphere; microaerobic = 42 ± 1°C in microaerobic atmosphere, 85% nitrogen, 10% carbon dioxide, 5% oxygen).
[b] N = Number of test portions.
[c] X = Number of positive test portions.
[d] POD_C = Probability of detection for candidate method confirmed positive outcomes.
[e] POD_R = Probability of detection for reference method confirmed positive outcomes.
[f] $dPOD_C$ = Difference between the candidate method and reference method POD values.
[g] 95% CI = If the confidence interval of a dPOD does not contain zero, then the difference is statistically significant at the 5% level.
[h] Trial performed by independent laboratory.

Table 6. Probability of detection calculations for ANSR for *Campylobacter* confirmed and reference method results after 24 h microaerobic or aerobic enrichment of naturally-contaminated samples (1)

Sample type	ANSR Incubation Condition[a]	N[b]	ANSR confirmed result			Reference method result			dPOD$_C$[f]	95% CI[g]
			X[c]	POD$_C$[d]	95% CI	X	POD$_R$[e]	95% CI		
Chicken Rinse Sample Set 1	Aerobic	20	14	0.70	0.48, 0.85	17	0.85	0.64, 0.95	-0.15	-0.39, 0.11
	Microaerobic	20	14	0.70	0.48, 0.85	17	0.85	0.64, 0.95	-0.15	-0.39, 0.11
Chicken Rinse Sample Set 2	Aerobic	20	15	0.75	0.53, 0.88	16	0.80	0.58, 0.92	-0.05	-0.30, 0.21
	Microaerobic	20	11	0.55	0.34, 0.74	11	0.55	0.34, 0.74	0.00	-0.28, 0.28
Chicken Rinse Sample Set 1[h]	Aerobic	20	5	0.25	0.11, 0.47	7	0.35	0.18, 0.57	-0.10	-0.36, 0.18
	Microaerobic	20	7	0.35	0.18, 0.57	6	0.30	0.15, 0.52	0.05	-0.23, 0.32
Chicken Rinse Sample Set 2[h]	Aerobic	20	6	0.30	0.15, 0.52	8	0.40	0.22, 0.61	-0.10	-0.36, 0.18
	Microaerobic	20	5	0.25	0.11, 0.47	5	0.25	0.11, 0.47	0.00	-0.26, 0.26
Turkey Sponge Sample Set 1	Aerobic	20	20	1.00	0.84, 1.00	20	1.00	0.84, 1.00	0.00	-0.16, 0.16
	Microaerobic	20	19	0.95	0.76, 1.00	20	1.00	0.84, 1.00	-0.05	-0.24, 0.12
Turkey Sponge Sample Set 2	Aerobic	20	7	0.35	0.18, 0.57	7	0.35	0.18, 0.57	0.00	-0.28, 0.28
	Microaerobic	20	5	0.25	0.11, 0.47	7	0.35	0.18, 0.57	-0.10	-0.36, 0.18

[a] The enrichment condition for the ANSR method (aerobic = 42 ± 1°C in normal atmosphere; microaerobic = 42 ±± 1°C in microaerobic atmosphere, 85% nitrogen, 10% carbon dioxide, 5% oxygen).
[b] N = Number of test portions.
[c] X = Number of positive test portions.
[d] POD$_C$ = Probability of detection for candidate method confirmed positive outcomes.
[e] POD$_R$ = Probability of detection for reference method confirmed positive outcomes.
[f] dPOD$_C$ = Difference between the candidate method and reference method POD values.
[g] 95% CI = If the confidence interval of a dPOD does not contain zero, then the difference is statistically significant at the 5% level.
[h] Trial performed by independent laboratory.

REFERENCES CITED

1. Viator, R., Alles, S., Lee, Q.N., Hosking, E., Zhange, L., Tolan, J., Meister, E., Tovar, E., Pinkava, L., Mozola, M., and Rice, J., Evaluation of the Neogen ANSR® for *Campylobacter* Method for Detection of Thermophilic *Campylobacter* spp. In Chicken Carcass Rinse and Turkey Sponge Samples , AOAC® *Performance Tested*^{SM} certification number 071601.
2. USDA-FSIS (2016) *Microbiology Laboratory Guidebook*, Chapter 41.04, Isolation and Identification of *Campylobacter jejuni/coli/lari* from Poultry Rinse, Sponge and Raw Product Samples (Accessed February, 2016) http://www.fsis.usda.gov/wps/wcm/connect/0273bc3d-2363-45b3-befb-1190c25f3c8b/MLG-41.pdf?MOD=AJPERES