

GeneQuence® *Campylobacter*

PRIOR TO STARTING THE TEST:

Note: Food Samples should be prepared and enriched, following the directions outlined in the GeneQuence *Campylobacter* spp. test kit insert.

UPON OPENING THE KIT:



Add 6 mL of **lysis reagent buffer** (bottle 1b) directly to **lysis reagent concentrate** (bottle 1a).



Mix **wash solution** (bottle 4) with 950 mL of distilled or deionized water.



STEP 1

For each sample and control, label a 12 x 75 mm glass tube with appropriate sample designation.

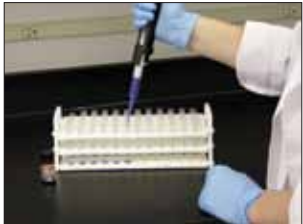


STEP 2

Place the appropriate number of microwells in the plate frame. Include first well for reagent blank, second well for negative control, and third well for positive control.

TEST PROCEDURE:

Add 0.4 mL of each of the sample or control to the appropriate tubes.



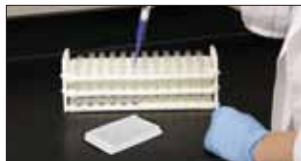
STEP 1

Add 0.1 mL of reconstituted **lysis reagent** (bottle 1a) to each tube.

Incubate the rack of tubes in the 54°C water bath or heater block for 30 minutes.



While the rack is incubating, refer to chart and mix **hybridization solution** (bottle 2) and **probe solution** (bottle 3) in a plastic or glass vial. Mix thoroughly.



STEP 2

Transfer 0.150 mL of each lysed sample, including the controls, to designated microwells.



STEP 3

Add 0.125 mL of **hybridization/probe mixture** to each microwell, with the exception of the reagent blank microwell.

Incubate the plate at room temperature for 60 minutes.



STEP 4

Wash the microwells 5 times.



STEP 5

Add 0.150 mL of **substrate chromagen** solution (bottle 5) to each microwell, including the blank microwell.

Incubate the plate at room temperature for 20 minutes.



STEP 6

Add 0.050 mL of **stop solution** (bottle 6) to each microwell, including the blank microwell.

Read absorbance at 450 nm using a plate strip reader.



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